

This article was downloaded by: [Malmo Hogskola]

On: 18 December 2011, At: 23:12

Publisher: Taylor & Francis

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



Journal of Asian Natural Products Research

Publication details, including instructions for authors and subscription information:

<http://www.tandfonline.com/loi/ganp20>

Three new flavonoid glycosides from *Urena lobata*

Lu Jia^a, You-Mei A^a, Lin-Lin Jing^a, Sheng-An Zhou^a & De-Yun Kong^b

^a School of Pharmacy, Zhengzhou University, Zhengzhou, 450001, China

^b Shanghai Institute of Pharmaceutical Industry, Shanghai, 200040, China

Available online: 05 Oct 2011

To cite this article: Lu Jia, You-Mei A, Lin-Lin Jing, Sheng-An Zhou & De-Yun Kong (2011): Three new flavonoid glycosides from *Urena lobata*, *Journal of Asian Natural Products Research*, 13:10, 907-914

To link to this article: <http://dx.doi.org/10.1080/10286020.2011.599802>

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: <http://www.tandfonline.com/page/terms-and-conditions>

This article may be used for research, teaching, and private study purposes. Any substantial or systematic reproduction, redistribution, reselling, loan, sub-licensing, systematic supply, or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae, and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand, or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

Three new flavonoid glycosides from *Urena lobata*

Lu Jia^a, You-Mei A^a, Lin-Lin Jing^a, Sheng-An Zhou^a and De-Yun Kong^{b*}

^aSchool of Pharmacy, Zhengzhou University, Zhengzhou 450001, China; ^bShanghai Institute of Pharmaceutical Industry, Shanghai 200040, China

(Received 7 February 2011; final version received 21 June 2011)

Three new flavonoid glycosides, kaempferol-3-*O*- β -D-apiofuranosyl(1 \rightarrow 2)- β -D-glucopyranosyl-7-*O*- α -L-rhamnopyranoside (**1**), kaempferol-4'-*O*- β -D-apiofuranosyl-3-*O*- β -D-glucopyranosyl-7-*O*- α -L-rhamnopyranoside (**2**), and 5,6,7,4'-tetrahydroxy-flavone-6-*O*- β -D-arabinopyranosyl-7-*O*- α -L-rhamnopyranoside (**3**), were isolated from the aerial parts of *Urena lobata* L., along with 10 known compounds (**4**–**13**). Their structures were determined based on spectroscopic methods including 1D and 2D NMR spectroscopy as well as HR-ESI-MS.

Keywords: Malvaceae; *Urena lobata* L; flavonoid glycoside

1. Introduction

As a traditional Chinese medicine, *Urena lobata* L. has been used to treat cold, fever, pain, or numbness caused by rheumatism, dysentery, edema, gonorrhea, leukorrhea, hematemesis, carbuncle, trauma bleeding, etc. In our earlier study on the chemical constituents of this plant, two new lignan compounds have been reported, i.e. ceplignan-4-*O*- β -D-glucopyranoside and 2,5-dihydroxy benzoic acid-7-(2,6-dimethyl-6-hydroxy-2,7-octadienoic acid)-anhydride-5-*O*- β -D-apiofuranosyl(1 \rightarrow 2)- β -D-glucopyranoside [1]. Further examination of this plant led to the isolation of three new flavonoid glycosides (**1**–**3**) and 10 known compounds (**4**–**13**). Herein, we report the isolation and the structural elucidation of the new compounds.

2. Results and discussion

Compound **1** was obtained as pale yellow amorphous powder with $[\alpha]_D^{26} - 6.9$. Its UV

absorption maxima at 267, 349 nm, etc., and positive HCl–Mg and Molish reactions indicated that compound **1** was a flavonoid glycoside. The molecular formula C₃₂H₃₈O₁₉ was determined from its quasi-molecular ion peak at m/z 749.1902 [M + Na]⁺ by its HR-ESI-MS, indicating 14 degrees of unsaturation. The ¹H NMR spectrum of **1** showed four aromatic protons of an AA'BB' system at δ 8.04/6.74 (each 2H, d, $J = 8.8$ Hz), and an AB type signal at δ 6.70 (1H, d, $J = 2.0$ Hz, H-8) and 6.40 (1H, d, $J = 2.0$ Hz, H-6). In addition, it showed three anomeric protons at δ 5.50 (1H, d, $J = 7.6$ Hz), 5.55 (1H, s), and 5.48 (1H, d, $J = 1.3$ Hz). Acid hydrolysis with trifluoroacetic acid afforded aglycone, glucose, rhamnose, and apiofuranose, which were identified by comparing with authentic sample on CO-HP-Si-TLC. The apiofuranosyl moiety can make complexation reaction with boric acid, therefore we identified its 2-hydroxy and 3-hydroxy as cis [2]. The stereochemistry of sugar units

*Corresponding author. Email: deyunk@yahoo.com.cn

was further confirmed by comparing their NMR spectral data with those in the literatures [3]. The ^{13}C NMR spectrum showed 32 carbon signals that were resolved as six methine aromatic carbons [C-2'/C-6' (δ_{C} 132.4), C-3'/C-5' (δ_{C} 118.6), C-6 (δ_{C} 100.3), C-8 (δ_{C} 95.2)], nine quaternary carbons (δ_{C} 107.5, 118.7, 134.1, 157.8, 157.8, 160.2, 163.0, 163.1, 179.2), six carbons for β -D-glucopyranosyl moiety (δ_{C} 62.6, 71.7, 78.2, 78.3, 79.1, 100.7), six carbons for α -L-rhamnopyranosyl moiety (δ_{C} 18.0, 71.6, 71.2, 72.1, 73.6, 99.8), and five carbons for β -D-apiofuranosyl moiety (δ_{C} 66.3, 75.6, 78.6, 80.9, 110.6) (Table 1). It was also confirmed by the HSQC and DEPT experiments. The ^1H and ^{13}C NMR spectra of **1** were almost identical to those of kaempferol-3-O- α -L-rhamnopyranoside-7-O-[α -D-apiofuranosyl-(1 \rightarrow 2)- β -D-glucopyranoside], except for the different sugar linkages [4]. The HMBC correlations from H-1'' (δ 5.50) to C-3 (δ 134.1), from H-1 to apiofuranosyl to C-2'' at δ 79.1, and from H-1''' (δ 5.55) to C-7 (δ 163.1) showed that β -D-apiofuranosyl-(1 \rightarrow 2)- β -D-glucopyranosyl moiety was attached to C-3, whereas the rhamnopyranosyl moiety was attached to C-7. Therefore, the structure of compound **1** was determined as kaempferol-3-O- β -D-apiofuranosyl(1 \rightarrow 2)- β -D-glucopyranosyl-7-O- α -L-rhamnopyranoside (Figure 1).

Compound **2** was isolated as yellow amorphous powder with $[\alpha]_{\text{D}}^{26} -2.3$ and gave positive reactions with HCl-Mg and Molish reagents, indicating that it was also a flavonoid glycoside. The UV spectrum displayed absorption maxima at 270, 356 nm, etc. The molecular formula $\text{C}_{32}\text{H}_{38}\text{O}_{19}$, with 14 degrees of unsaturation, was deduced from its HR-ESI-MS, which showed a quasi-molecular ion peak at m/z 749.1901 $[\text{M} + \text{Na}]^+$, consistent with its NMR spectral data.

The ^1H and ^{13}C NMR spectra of **2** resembled those of **1**, except for the sugar linkage (Table 1). The locations of the sugar linkage were concluded to be C(3)-C(1''), C(7)-C(1'''), and C(4')-C(1''')

based on the HMBC experiment. The HMBC correlation of **2** from H-1'' (δ 5.52) to C-3 (δ 134.9) confirmed the attachment of β -D-glucosyl unit to the aglycone, and a down-field shift of C-2 (δ 157.8) was the other evidence for the linkage of β -D-glucose. The position of the rhamnose unit was confirmed by correlations from H-1''' (δ 5.56) to C-7 (δ 163.2), and a down-field shift of H-6 (δ 6.45) and H-8 (δ 6.75) could be seen due to the glucosidation of C-7. Similarly, the position of β -D-apiofuranosyl unit was confirmed by correlations from H-1'''' (δ 5.45) to C-4' (163.1). A down-field shift of C-1' (δ 122.7) and H-3'/5' (δ 6.91) was the other evidence for the location of β -D-apiofuranosyl moiety. Therefore, compound **2** was elucidated as kaempferol-4'-O- β -D-apiofuranosyl-3-O- β -D-glucopyranosyl-7-O- α -L-rhamnopyranoside (Figure 1).

Compound **3** was obtained as pale yellow amorphous powder. It showed positive reactions with HCl-Mg and Molish reagents, indicating that it was a flavonoid glycoside. The molecular formula was determined as $\text{C}_{26}\text{H}_{28}\text{O}_{14}$ based on the pseudo-molecular ion at m/z 587.1371 $[\text{M} + \text{Na}]^+$.

The ^1H NMR spectrum of **3** showed aromatic protons at δ 7.98/6.93 (each 2H, d, $J = 8.8$ Hz), 6.99 (1H, s, H-8), and 6.86 (1H, s, H-3). The anomeric protons of β -D-arabinopyranosyl and α -L-rhamnopyranosyl moieties appeared at δ 5.09 (1H, d, $J = 4.0$ Hz, H-1'') and 5.54 (1H, d, $J = 1.6$ Hz, H-1'''), and other characteristic proton signals for β -D-arabinopyranose and α -L-rhamnopyranose were also displayed in its ^1H NMR spectrum [5]. Acid hydrolysis with trifluoroacetic acid afforded arabinose and rhamnose, which were identified by comparing with authentic sample on CO-HP-Si-TLC. The ^{13}C NMR spectral data showed 26 carbon signals, including 15 carbons for the aglycone, five carbons for β -D-arabinopyranosyl moiety (δ_{C} 62.8, 69.9, 69.6, 70.4, 102.2), and six carbons for the

Table 1. ^1H , ^{13}C NMR, and HMBC spectroscopic data for compounds **1** and **2** (measured in DMSO- d_6).

No.	1				2			
	$\delta^{13}\text{C}$	$\delta^1\text{H}$ (J in Hz)	DEPT	HMBC	$\delta^{13}\text{C}$	$\delta^1\text{H}$ (J in Hz)	DEPT	HMBC
Flavonoids								
2	157.8	—	C	—	157.8	—	C	—
3	134.1	—	C	—	134.9	—	C	—
4	179.2	—	C	—	179.5	—	C	—
5	160.2	—	C	—	161.4	—	C	—
6	100.3	6.40 d ($J = 2.0$)	CH	C-7, C-8, C-10	100.5	6.45 s	CH	C-7, C-8, C-10
7	163.1	—	C	—	163.2	—	C	—
8	95.2	6.70 d ($J = 2.0$)	CH	C-6, C-10	95.4	6.75 s	CH	C-6, C-10
9	157.8	—	C	—	159.1	—	C	—
10	107.5	—	C	—	107.5	—	C	—
1'	118.7	—	C	—	122.7	—	C	—
2'	132.4	8.04 d ($J = 8.8$)	CH	C-2, C-1', C-4'	132.2	8.08 d ($J = 8.8$)	CH	C-2, C-1', C-4'
3'	118.6	6.74 d ($J = 8.8$)	CH	C-1', C-2'	116.2	6.91 d ($J = 8.8$)	CH	C-1', C-2'
4'	163.0	—	C	—	163.1	—	C	—
5'	118.6	6.74 d ($J = 8.8$)	CH	C-1', C-6'	116.2	6.91 d ($J = 8.8$)	CH	C-1', C-6'
6'	132.4	8.04 d ($J = 8.8$)	CH	C-2, C-1', C-4'	132.2	8.08 d ($J = 8.8$)	CH	C-2, C-1', C-4'
Glucosyl								
1''	100.7	5.50 d ($J = 7.6$)	CH	C-3	100.5	5.52 d ($J = 7.2$)	CH	C-3
2''	79.1	3.66 m	CH	—	78.0	3.64 m	CH	—
3''	78.2	4.04 m	CH	—	78.1	4.05 m	CH	—
4''	71.7	4.00 m	CH	—	71.5	4.01 m	CH	—
5''	78.3	3.59 m	CH	—	79.2	3.60 m	CH	—
6''	62.6	3.65, 3.44 m	CH ₂	—	62.4	3.66, 3.46 m	CH ₂	—
Rhamnosyl								
1'''	99.8	5.55 s	CH	C-7, C-5'''	99.6	5.56 s	CH	C-7
2'''	72.1	3.82 m	CH	—	71.9	3.85 m	CH	—
3'''	71.6	3.59 m	CH	—	71.1	3.82 m	CH	—
4'''	73.6	3.45 m	CH	—	73.5	3.44 m	CH	—
5'''	71.2	3.60 m	CH	—	70.9	3.61 m	CH	—
6'''	18.0	1.24 d ($J = 6.1$)	CH ₃	C-4''', C-5'''	18.0	1.24 d ($J = 6.4$)	CH ₃	—

Table 1 – continued

No.	1				2			
	$\delta^{13}\text{C}$	$\delta^1\text{H}$ (J in Hz)	DEPT	HMBC	$\delta^{13}\text{C}$	$\delta^1\text{H}$ (J in Hz)	DEPT	HMBC
Apiofuranosyl								
1	110.6	5.48 d ($J = 1.3$)	CH	C-2 ^{''} , C-3 ^{'''}	110.6	5.45 s	CH	C-4'
2	78.6	4.06 m	CH	C-4 ^{'''} , C-5 ^{'''}	79.2	4.04 m	CH	
3	80.9	–	C		80.8	–	C	
4	75.6	3.71, 4.08 m	CH ₂	C-1 ^{'''} , C-2 ^{'''}	75.3	3.71, 4.05 m	CH ₂	
5	66.3	3.74 d ($J = 11.2$)	CH ₂	C-2 ^{'''} , C-3 ^{'''}	66.0	3.73 d ($J = 10.0$)	CH ₂	

α -L-rhamnopyranosyl moiety (δ_{C} 18.2, 69.7, 71.5, 71.9, 73.6, 99.5) (Table 2), which was also supported by the HSQC and DEPT experiments. The HMBC correlation of **3** from H-1^{''} (δ 5.09) to C-6 (δ 128.7) confirmed the attachment of an arabinose unit to the aglycone, and the position of the rhamnose unit was confirmed in a similar manner by the correlation from H-1^{'''} (δ 5.54) to C-7 (δ 155.3). Therefore, compound **3** was determined as 5,6,7,4'-tetra-hydroxyflavone-6-*O*- β -D-arabinopyranosyl-7-*O*- α -L-rhamnopyranoside (Figure 1).

Compounds **4**–**13** were identified as 5,6,7,4'-tetrahydroxy-flavone-6-*O*- β -D-xylopyranosyl-7-*O*- α -L-rhamnopyranoside (**4**) [6], kaempferol-7-*O*- β -D-glucopyranosyl (1 \rightarrow 3)- α -L-rhamnopyranoside (**5**) [7], kaempferol-3-*O*- β -D-glucopyranosyl-7-*O*- α -L-rhamnopyranoside (**6**) [8], kaempferol-3-*O*- β -D-glucopyranoside (**7**) [9], 6,8-dihydroxykaempferol-3-*O*- β -D-glucopyranoside (**8**) [10], kaempferol-4'-*O*- β -D-glucopyranoside (**9**) [11], kaempferol-7-*O*- α -L-rhamnopyranoside (**10**) [12], kaempferol-7-*O*- α -L-rhamnopyranoside-4'-*O*- β -D-glucopyranoside (**11**) [13], kaempferol-3-*O*-glucopyranosyl(1 \rightarrow 3)- β -D-glucopyranoside (**12**) [14], and kaempferol-3-*O*- β -robinobioside (**13**) [15] by comparing their spectroscopic data with those reported in the literatures.

3. Experimental

3.1 General experimental procedures

Optical rotations were determined on a Perkin–Elmer 341 polarimeter (Perkin–Elmer Co., Norwalk, USA). UV spectral data were acquired on a JASCO P-1030 spectrophotometer (JASCO Corporation, Tokyo, Japan). ¹H and ¹³C NMR spectra were measured using a Bruker AVANCE DPX-400 spectrometer (Bruker Corporation, Fällanden, Switzerland). Mass spectra were measured using a Waters Q-TOF micro mass spectrometer (Waters Corporation, Milford, MA, USA). Column

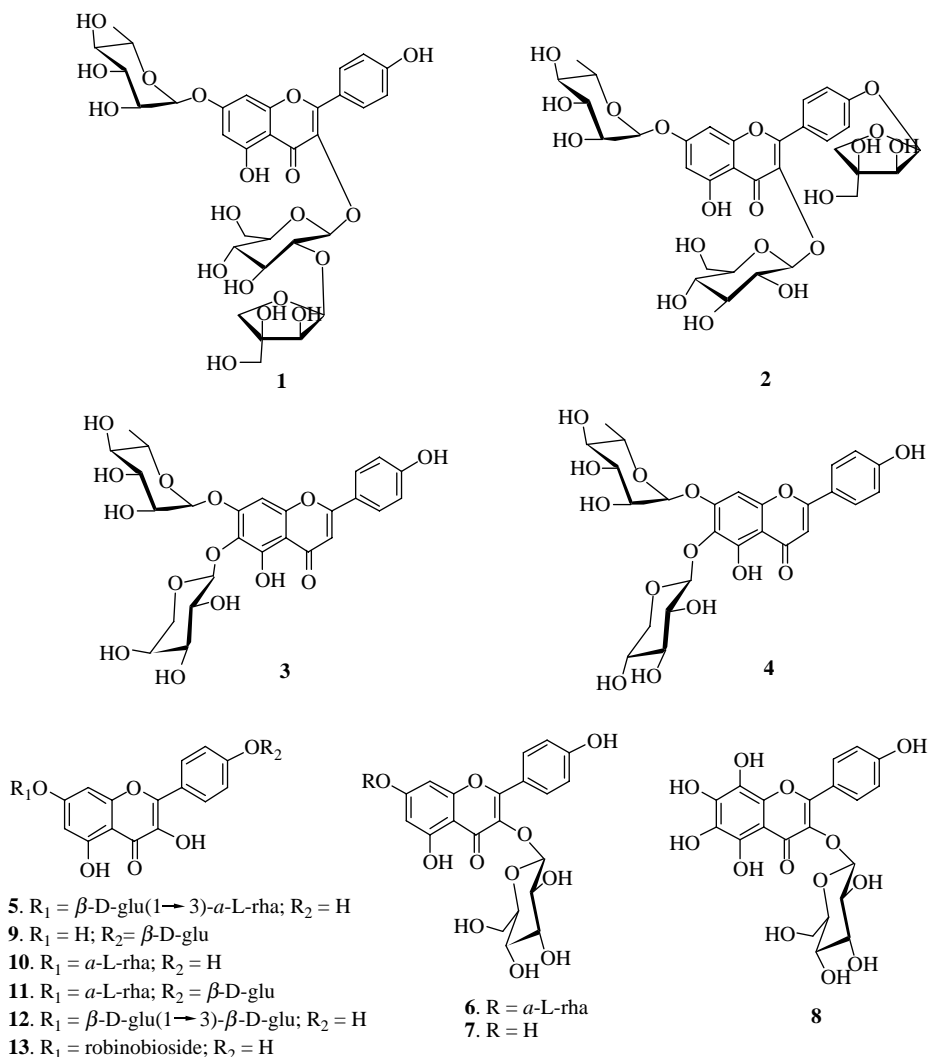


Figure 1. Structures of compounds 1–13.

chromatography (CC) was carried out on Sephadex LH-20 (Pharmacia Corporation, New York, NY, USA), silica gel 100–200 mesh (Qingdao Haiyang Chemical Co., Ltd, Qingdao, China). Silica gel GF₂₅₄ (Qingdao Haiyang Chemical Co.) was used for TLC.

3.2 Plant material

The aerial parts of *U. lobata* were collected from Quanzhou city of Fujian

Province, China, and identified by Dr Tong Wu from Shanghai Institute of Pharmaceutical Industry. A voucher specimen (No. 20060823) is deposited in Shanghai Institute of Pharmaceutical Industry.

3.3 Extraction and isolation

The aerial parts of *U. lobata* (30 kg) were crushed and extracted with 95% EtOH (10 × 3 liters) under reflux. The extract was concentrated *in vacuo* to give a

Table 2. ^1H , ^{13}C NMR and HMBC spectroscopic data for compound **3** (measured in $\text{MeOH}-d_4$).

		3							
No.	$\delta^{13}\text{C}$	$\delta^1\text{H}$ (HSQC) (J in Hz)	DEPT	HMBC	No.	$\delta^{13}\text{C}$	$\delta^1\text{H}$ (HSQC) (J in Hz)	DEPT	HMBC
Flavonoids									
2	164.6	-	C	-	6'	128.9	7.98 d ($J = 8.8$)	CH	C-2, C-4'
3	102.2	6.86 s	CH	C-2, C-1', C-10	Arabinosyl			CH	C-6
4	182.5	-	C	-	1''	102.2	5.09 d ($J = 4.0$)	CH	-
5	152.7	-	C	-	2''	69.9	3.91 m	CH	-
6	128.7	-	C	-	3''	69.9	3.91 m	CH	-
7	155.3	-	C	-	4''	70.4	3.48 m	CH	-
8	94.5	7.01 s	CH	C-6, C-7, C-9, C-10	5''	62.8	3.82, 3.31 m	CH_2	-
9	152.5	-	C	-	Rhamnosyl			CH	C-7
10	105.8	-	C	-	1'''	99.5	5.54 d ($J = 1.6$)	CH	-
1'	121.3	-	C	-	2'''	71.9	3.33 m	CH	-
2'	128.9	7.98 d ($J = 8.8$)	CH	C-2, C-4'	3'''	71.5	3.60 m	CH	-
3'	116.2	6.93 d ($J = 8.8$)	CH	C-1', C-4'	4'''	73.6	3.32 m	CH	-
4'	161.6	-	C	-	5'''	69.7	3.43 m	CH	-
5'	116.2	6.93 d ($J = 8.8$)	CH	C-1', C-4'	6'''	18.2	1.17 d ($J = 6.1$)	CH_3	-

residue (1.75 kg), which was dissolved in water (4 liters), and then partitioned with petroleum ether (3 × 2 liters), chloroform (3 × 2 liters), ethyl acetate (3 × 2 liters), and *n*-butanol (3 × 2 liters), respectively. The *n*-butanol fraction (345 g) was dissolved in water and passed through a D₁₀₁ macroporous adsorption resin column (10 × 120 cm) eluted with H₂O:EtOH (100:0, 80:20, 60:40, 40:60, 20:80, 0:100, v/v) to yield seven fractions (Fr. 1–7). Fr. 2 (15.3 g) was chromatographed on a Sephadex LH-20 column with MeOH as eluent to give three subfractions. Subfraction 2 (5.2 g) was chromatographed on silica gel column (with a gradient of EtOAc–MeOH–H₂O from 30:2:1 to 5:2:1) to give **3** (10 mg), **4** (18 mg), **5** (6 mg), and **7** (5 mg). Fraction 4 (23.8 g) was applied to Sephadex LH-20 CC with MeOH as eluent to give six subfractions 4A–4F. Fraction 4B (3.6 g) was subjected to both Sephadex LH-20 CC (MeOH) and silica gel CC (with a gradient of EtOAc–MeOH–H₂O from 30:2:1 to 10:2:1), repeatedly, to give **1** (3 mg), **2** (6 mg), and **6** (12 mg). Fraction 4C (1.9 g) was repeatedly applied to Sephadex LH-20 CC (MeOH) and silica gel CC (with a gradient of EtOAc–MeOH from 20:1 to 3:1) to give **8** (3 mg), **9** (4 mg), **12** (1.2 mg), and **13** (1.7 mg). Fraction 4D (3.8 g) was repeatedly applied to Sephadex LH-20 CC (MeOH) and silica gel column (with a gradient of EtOAc–MeOH from 20:1 to 3:1) to give **10** (1.6 mg) and **11** (3 mg).

3.3.1 *Kaempferol-3-O-β-D-apiofuranosyl (1 → 2)-β-D-glucopyranosyl-7-O-α-L-rhamnopyranoside*

C₃₂H₃₈O₁₉; pale yellow amorphous powder; $[\alpha]_D^{26} - 6.9$ ($c = 0.61$, CH₃OH); IR ν_{\max} (cm⁻¹): 3385, 1663, 1592, 1500; UV λ_{\max} (CH₃OH, nm): 267, 349, etc. ¹H and ¹³C NMR spectral data see Table 1; HR-ESI-MS: m/z 749.1902 [M + Na]⁺ (calcd for C₃₂H₃₈O₁₉Na, 749.1905).

3.3.2 *Kaempferol-4'-O-β-D-apiofuranosyl-3-O-β-D-glucopyranosyl-7-O-α-L-rhamnopyranoside*

C₃₂H₃₈O₁₉; pale yellow amorphous powder; $[\alpha]_D^{26} - 2.3$ ($c = 0.31$, DMSO); IR ν_{\max} (cm⁻¹): 3384, 1665, 1593, 1500; UV λ_{\max} (DMSO, nm): 270, 356, etc. ¹H and ¹³C NMR spectral data see Table 1; HR-ESI-MS: m/z 749.1901 [M + Na]⁺ (calcd for C₃₂H₃₈O₁₉Na, 749.1905).

3.3.3 *5,6,7,4'-Tetrahydroxyflavone-6-O-β-D-arabinopyranosyl-7-O-α-L-rhamnopyranoside*

C₂₆H₂₈O₁₄; pale yellow amorphous powder; $[\alpha]_{589}^{24.3} - 54$ ($c = 0.17$, CH₃OH); IR ν_{\max} (cm⁻¹): 3400, 1660, 1607, 1499; UV λ_{\max} (DMSO, nm): 275, 332, etc. ¹H and ¹³C NMR spectral data see Table 2; HR-ESI-MS: m/z 587.1371 [M + Na]⁺ (calcd for C₂₆H₂₈O₁₄Na, 587.1377).

References

- [1] L. Jia, Y.F. Bi, L.L. Jing, S.A. Zhou, and D.Y. Kong, *J. Asian Nat. Prod. Res.* **12**, 962 (2010).
- [2] J. Huang, M.L. Tu, and J.X. Xie, *Acta Pharm. Sin.* **22**, 264 (1987).
- [3] L.P. Anna, V. Attilio, P. Siro, D.S. Francesco, and R. Luca, *Food Chem.* **100**, 344 (2007).
- [4] Y.H. Chen, F.R. Chang, Y.J. Lin, L.S. Wang, J.F. Chen, Y.C. Wu, and M.J. Wu, *Food Chem.* **105**, 48 (2007).
- [5] D.Q. Yu and J.S. Yang, *Handbook of Analytical Chemistry* (Chemical Industry Press, Beijing, 1999), Vol. 7, p. 492.
- [6] A.G. Ramachandran Nair, T.R. Seetharaman, B. Voirin, and J. Favre-Bonvin, *Phytochemistry* **25**, 768 (1986).
- [7] M. Du and J.M. Xie, *Phytochemistry* **38**, 809 (1995).
- [8] R. Nakabayashi, M. Kusano, M. Kobayashi, T. Tohge, K. Yonekura-Sakakibara, N. Kogure, M. Yamazaki, M. Kitajima, and K. Saito, *Phytochemistry* **70**, 1017 (2009).
- [9] A. Palida, R.S. Mi, Y.Y. Cong, D. Yiba, and X.W. Wang, *West Chin. J. Pharm. Sci.* **22**, 489 (2007).

- [10] M.A. Andreo, K.V. Ballesteros, C.A. Hiruma Lima, L.R. Machado da Rocha, A.R. Souza Brito, and W. Vilegas, *J. Ethnopharmacol.* **107**, 431 (2005).
- [11] T. Scheer and M. Wichtl, *Planta Med.* **53**, 573 (1987).
- [12] F. Shaheen, L. Ali, S. Ali, N. Erdemoglu, and B. Sener, *Chem. Nat. Comp.* **45**, 346 (2009).
- [13] I. Filippo, *Phytochemistry* **47**, 911 (1998).
- [14] T. Morikawa, L.B. Wang, S. Nakamura, K. Ninomiya, E. Yokoyama, H. Matsuda, O. Muraoka, L.J. Wu, and M. Yoshikawa, *Chem. Pharm. Bull.* **57**, 361 (2009).
- [15] N.Y. Yang, J.A. Duan, and P. Li, *Chin. Pharm. J.* **21**, 1621 (2006).